Novel Application of Laboratory

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Instrumentation Characterizes Mass Settling

Dynamics of Oil-Mineral Aggregates (OMAs)

and Oil-Mineral-Microbial Interactions

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ABSTRACT

It is reasonable to assume that microbes played an important role in determining the eventual fate of oil spilled during the 2010 Deepwater Horizon disaster, given that microbial activities in the Gulf of Mexico are significant and diverse. However, critical gaps exist in our knowledge of how microbes influence the biodegradation and accumulation of petroleum in the water column and in marine sediments of the deep ocean and the shelf. Ultimately, this limited understanding impedes the ability to forecast the fate of future oil spills, specifically the capacity of numerical models to simulate the transport and fate of petroleum under a variety of conditions and regimes.

By synthesizing recent model developments and results from field- and laboratorybased microbial studies, the Consortium for Simulation of Oil-Microbial Interactions in the Ocean (CSOMIO) investigates (a) how microbial biodegradation influences accumulation of petroleum in the water column and in marine sediments and (b) how biodegradation can be influenced by environmental conditions and impact forecasts of potential future oil spills.

Keywords:

Laboratory Flocculation **Experiments**

ritical to oil-mineral-microbial interactions is a process whereby cohesive sediment particles do not behave as individual, dispersed particles but instead tend to stick together. This process is known as flocculation, and the resultant floc sizes and settling velocity are much greater than those of the individual constituent particles, but their overall floc effective density is less (e.g., Dyer & Manning, 1999; Manning & Dyer, 1999). When oil droplets are contained by

55 marine snows, oil sedimentation can 56 occur and provide an unexpected 57 pathway in the oil budget calculation 58 (Daly et al., 2016; Muschenheim & 59 Lee, 2002; Passow & Ziervogel, 60 2016). A novel high-resolution floc 61 video instrument originally designed 62 to determine the spectral characteris-63 tics of flocculating cohesive sediments 64 has, for the first time, been applied to 65 study floc size distribution and set-66 tling dynamics of oil-mineral aggre-67 gates (OMAs). The results of this 68 study inform the development of 69 efficient and accurate algorithms for 70 simulating the formation and settling flocs of cohesive sediment and/or 71 of these flocs.

As part of the Consortium for Simulation of Oil-Microbial Interactions in the Ocean (CSOMIO), a series of laboratory flocculation experiments with seawater, crude oil, and cohesive sediment mixtures (mineral clay and artificial extracellular polymeric substances) have been conducted at the Center for Applied Coastal Research, University of Delaware, using the LabSFLOC-2 (the second generation of the LabSFLOC [Laboratory Spectral Flocculation Characteristics instrument; Manning, 2015], developed by Manning, 2006). In these experiments, the LabSFLOC-2 instrument, a digital video microscope and

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processing package, makes it possible to obtain high-quality floc population data (e.g., individual floc size, settling velocity, density, mass), as well as supplementary individual floc information including floc porosity, floc mass, fractal dimension, floc shape, and mass settling flux. Manning et al. (2010) and Manning et al. (2017) provide further details of the floc acquisition procedures and postprocessing computations, respectively. LabSFLOC-2 provides data for many important aspects of flocculation. These floc data are necessary to comprehensively assess settling dynamics and to improve the 136 magnification, F4, macro lens. parameterization (Manning & Dyer, 137 OMAs.

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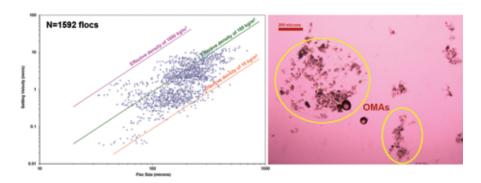
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Laboratory Experiments Utilizing the LabSFLOC-2 Instrument

2.0-MP Grasshopper monochrome 155 subsequent floc settling process. digital video camera to optically ob- 156 resolution) telecentric (maximum 167 fluid head of 50 mm, which results in

FIGURE 1

The LabSFLOC-2 setup on the desk beside the stir jar system for real-time samplings (photo provided by Prof. A. J. Manning).



and characterize oil-mineral-microbial 135 pixel distortion of 0.6%), 0.66 (1:1.5)

A suspension containing oil-mineral-2007; Soulsby et al., 2013) and calibra- 138 microbial flocs is initially introduced tion (Baugh & Manning, 2007) of 139 to the LabSFLOC-2 column, while a numerical models. Additionally, the 140 suspension is extracted from the digital microscope images help us better 141 jar fluid using a specially modified understand the visible floc structure of 142 Serological TD-EX 20°C 50-ml 143 maximum-capacity sterile pipette. 144 This process has proved to be mini-145 mally intrusive for flocs, relying only 146 upon settling due to gravity and thus 147 avoiding the need for additional 148 fluid or turbulence transfer. The Mass settling dynamics of oil- 149 Lab SFLOC-2 instrumentation is mineral flocs are observed using the 150 located close and adjacent to the stir LabSFLOC-2 system (Figure 1), 151 jar system, as this minimizes the time which measures an entire floc popula- 152 needed to transfer floc samples to the tion for each sample being assessed. 153 LabSFLOC-2 settling chamber and LabSFLOC-2 utilizes a low intrusive 154 any potential disruption during the

The camera views through an aperserve individual flocs (e.g., Manning 157 ture in the settling column wall at a & Dyer, 2002) as they settle in a 158 depth of 230 mm below the column 350 mm high × 100 mm square 159 water surface. It records all settling Perspex settling column. The video 160 flocs/particles in the center of the camera, positioned nominally 75 mm 161 column, which pass within a 1-mm above the base of the column, views 162 focal depth of field, 45 mm (focal all particles in the center of the column 163 length) from the camera lens. The that pass within a 1-mm depth of field, 164 total image size is nominally 6 mm 45 mm from the Sill TZM 1560 high- 165 high and 8 mm wide. During sammagnification (nominal 5-µm pixel 166 pling, a pipette is filled to produce a a video image control sample volume nominally of 400 mm³ (1-mm image depth and 6-mm nominal video image width, with a nominal 50-mm high suspension extracted with a modified pipette). This control volume permits the LabSFLOC-2 calculated total floc mass to be accurately massbalanced with the nominal suspended particulate matter concentration utilized in the jar test under examination. The LabSFLOC-2 camera can view particles as small as 5 µm and as large as 8 mm. Settling velocities ranging from 0.01 to 45 mm·s⁻¹ can be measured by the LabSFLOC-2, and the system can operate within floc suspended particulate matter concentrations of a few milligrams per liter, with a practical upper operating limit of $\sim 200 \text{ g} \cdot \text{l}^{-1}$.

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Settling flocs are viewed as silhouettes (to reduce image smearing) resulting from a 43 × 35 mm, homogeneous blue (470 nm), back-illumination LED panel located at the rear of the settling column. The digital floc images are captured as non-Codec compressed AVI files at a frame rate of 7.5 Hz (one frame is 0.04 s), at a resolution of $1,600 \times 1,200$ pixels, with an individual pixel nominally representing 5 µm (confirmed by independent

FIGURE 2

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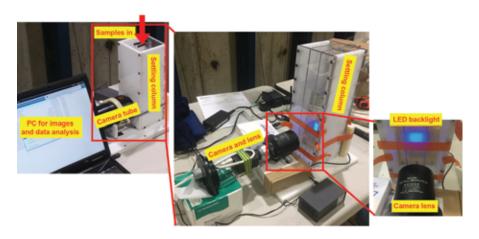
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Sample from an oil-bentonite case. The left plot shows the floc size and settling velocity scatters of each calculated floc. The three diagonal lines present contours of Stokes settling velocity calculated with a constant effective density (i.e., floc bulk density minus water density): pink = 1,600 kg·m⁻³ (equivalent to a quartz particle), green = 160 kg·m⁻³, and red = 16 kg·m⁻³. The right image is the generated OMAs as seen by the digital microscope camera (approximately ×40).



internal hard drive.

(e.g., Figure 2, right) but also reveals 234 density (16–160 kg·m⁻³) region. all other essential quantitative floc properties. The uncompressed images are then analyzed with MATLAB soft- 235 Summary ware routines. During postprocessing, 236

calibration), connected and streamed 229 30 and 700 µm, and settling velocito a laptop PC, and recorded on the 230 ties spanned 0.3-10 mm·s⁻¹. The 231 plot (Figure 2, left) shows a signifi-The present system not only pro- 232 cant portion of the floc population duces visible floc individual images 233 clusters within the low-effective

In the first attempt to apply the the HR Wallingford Ltd. DigiFloc 237 LabSFLOC-2 system in an oil-mineral software version 1.0 (Benson & 238 flocculation study, we have com-Manning, 2013) and JavaScript can 239 bined state-of-the-art technologies/ be used to semiautomatically process 240 instruments in order to expand our the digital recording image stack to 241 knowledge of oil-sediment-microbial obtain floc size and settling velocity 242 interactions and the vertical transport spectra (e.g., Figure 2, left for oil- 243 of oil. The preliminary laboratory bentonite flocs). A modified version 244 experiments demonstrate that these of Stokes' law (Stokes, 1851) permits 245 systems can be used to produce and an accurate estimate of individual floc 246 characterize mass settling dynamics of effective density (Manning et al., 247 OMAs. Future experiments will use 2013), which can then be utilized to 248 different oil, sediment, and microbial calculate floc mass. In the oil-bentonite 249 characteristics and turbulence levels. sample, resultant floc sizes (nominally 250 Statistical data on settling dynamics mass-balanced to a suspended par- 251 provided by LabSFLOC-2 will allow ticulate matter concentration of 252 for a systematic analysis of the role 1,000-mg·l⁻¹ bentonite and 1 ml of 253 that each factor plays in determining Texas crude oil) ranged between 254 the resultant settling dynamics. Moving forward, these technologies have the potential for applications to a carefully designed test matrix in order to calibrate a given modeling framework for oil-sediment-microbial interactions.

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